## Characterization of the anti-HIV-1 activity of DABOs.

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DABOs are specific inhibitors of HIV-1 targeted at the reverse transcriptase. From the chemical point of the view they are 6-benzyldihydrooxopyrimidines substituted at the position 2 with either alkoxy or alkylthio side chains.

DABOs potently and selectively inhibit in vitro the replication of several HIV-1 laboratory strains and clinical isolates in a variety of T and monocytic cells. Effective concentrations (EC<sub>50</sub>) are in the range 0.3- $1.0~\mu\text{M}$ , whereas doses up to  $300~\mu\text{M}$  are not cytotoxic. No activity against HIV-2 can be demonstrated at doses up to  $100~\mu\text{M}$ .

In enzyme assays DABOs inhibit HIV-1 recombinant RT at doses comparable to those inhibiting virus multiplication in cell cultures; their kinetic pattern of inhibition shows that they do not compete with natural substrates.

Highly resistant mutants to the most potent DABOs can be selected after 3 passages in the presence of high drug doses (50  $\mu$ M). Mutants are sensitive to AZT, ddI or ddC and partially cross-resistant to nevirapine and E-BPU-S (2 and 10 fold increase in EC<sub>50</sub>, respectively). The identification of the mutation responsible for DABO resistance is currently under investigation.

Combinations of DABOs with AZT or ddI result in additive/slightly synergistic anti-HIV-1 effects.

Long-term treatments of acutely infected MT-4 cells with combinations of a DABO derivative (2.5-10  $\mu$ M) with AZT (0.08-0.32  $\mu$ M) result in no viral breakthrough after 45 days. On the contrary virus breakthrough can be observed when DABO (10  $\mu$ M) is combined with ddI (up to 80  $\mu$ M) or ddC (up to 24  $\mu$ M).

We are currently investigating whether integrated provirus is present in there surviving HIV-infected, drug-treated cultures.

This work was supported in part by ISS grants n. 9204-68 and 9204-05.

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## Anti-HIV Activity of Bovine Seminal RNase.

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BS-RNasc (bovine seminal RNase), an enzyme acting both on single- and double-stranded RNA, is a versatile biological effector with special biological actions that include a potent anticancer activity both *in vitro* and *in vivo* (D'Alessio 1993, Trends Cell Biol. 3, 106-109).

Recently, BS-RNase has been shown to inhibit the HIV-1 replication in H9 cells at non-cytotoxic concentrations (10<sup>-7</sup> M). Other homologous RNases, such as RNase A (the well known model protein devoid of any special biological actions) and EDN (an RNase from human eosinophil granules with a selective cytotoxicity for certain neurons), have shown no anti-HIV-1 activity. On the contrary, onconase (an RNase with anticancer activity (Mikulski et al., 1990, J.Natl.Cancer Inst. 82, 151-152) from amphibian eggs) has been found to potently inhibit the HIV-1 replication.

We tested RNase A (as a reference compound) and BS-RNase for cytotoxicity and anti-HIV-1 activity in MT-4 and C8166 cells. In both cell lines, RNase A was non-cytotoxic at doses up to  $7 \times 10^{-5}$  M. On the contrary,  $CC_{50}$ s of BS-RNase for cell growth were  $10^{-7}$  M and  $4 \times 10^{-7}$  M in MT-4 and C8166, respectively.

When tested in acutely infected cells, RNase A resulted always ineffective, whereas the behaviour of BS-RNase was different according to the virus/cell system used. When evaluated in MT-4 cells, BS-RNase was ineffective in preventing the HIV-1-induced CPE but inhibited virus multiplication, as evaluated by p24 assay, with an EC $_{50}$  of  $5\times10^{-8}$  M. In acutely infected C8166 cells, and in C8166-H9/III $_{\rm B}$  cocultures, BS-RNase was effective in reducing both syncytium formation and p24 levels.

This work was supported by ISS grants n. 9204-68 and 9204-47.